

# Protein Loop Kinematics

A protein loop (or fragment)  $L$  is a sequence of  $p > 3$  consecutive residues in a protein  $P$ , such that none of the two termini of  $L$  is also a terminus of  $P$ . We number the residues of  $L$  from 1 to  $p$ , starting at the N terminus. We model the kinematics of the backbone of  $L$  as a serial linkage whose DOFs are the  $n = 2p$  dihedral angles  $\phi_i$  and  $\psi_i$  around the bonds N–C $\alpha$  and C $\alpha$ –C, in residues  $i = 1, \dots, p$ . The rest of the protein, denoted by  $P \setminus L$ , is assumed rigid. We let  $L_B$  denote the backbone of  $L$ . It includes the C $\beta$  and O atoms respectively bonded to the C $\alpha$  and C atoms in the backbone, as their positions are uniquely determined by the dihedral angles  $\phi_i$  and  $\psi_i$ .

To simplify our presentation, we will use the above model throughout. However, our methods are more general. They allow a loop to start at either an N or C $\alpha$  atom and end at either a C $\alpha$  or C atom. Furthermore, they do not require all  $\phi$  and  $\psi$  angles in  $L_B$  to be variable. Some may have fixed input values. We only require the loop to contain at least three amino-acids, consecutive or not, with variable  $\phi$  and  $\psi$  angles. This condition is needed for the application of the analytical IK method described in [Coutsias *et al.*, 2004].

We attach a Cartesian coordinate frame  $\Omega_1$  to the N terminus of  $L$  and another frame  $\Omega_2$  to its C terminus. When  $L_B$  is connected to the rest of the protein, i.e., when it adopts a *closed* conformation, the pose (position and orientation) of  $\Omega_2$  relative to  $\Omega_1$  is fixed. We denote this pose by  $\Pi_g$ . However, if we arbitrarily pick the values of  $\phi_i$  and  $\psi_i$ ,  $i = 1$  to  $p$ , then in general we get an *open* configuration of  $L_B$ , where the pose of  $\Omega_2$  differs from  $\Pi_g$ . The set  $\mathbf{Q}$  of all open and closed conformations of  $L_B$  is a manifold of dimensionality  $n = 2p$ . The subset  $\mathbf{Q}_{\text{closed}}$  of closed conformations is a submanifold of  $\mathbf{Q}$  of dimensionality  $n - 6$  for almost any  $\Pi_g$ ; it is sometimes called the “self-motion manifold” of  $L_B$ . When  $n = 6$ ,  $\mathbf{Q}_{\text{closed}}$  has zero dimensionality; it then contains at most 16 isolated closed conformations [Coutsias *et al.*, 2004, Liu *et al.*, 2006]. Let  $\Pi(q)$  denote the pose of  $\Omega_2$  relative to  $\Omega_1$  when the conformation of  $L_B$  is  $q \in \mathbf{Q}$ . The function  $\Pi$  and its inverse  $\Pi^{-1}$  are the “forward” and “inverse” kinematics map of  $L_B$ , respectively. We have:  $q \in \mathbf{Q}_{\text{closed}} \Leftrightarrow \Pi(q) = \Pi_g$ .

A conformation of  $L_B$  is *clash-free* if and only if no two atoms, one in  $L_B$ , the other in  $L_B$  or  $P \setminus L$ , are such that their centers are closer than  $\varepsilon$  times the sum of their van de Waals radii, where  $\varepsilon$  is a constant in  $(0, 1)$ . In our software,  $\varepsilon$  is an adjustable parameter, usually set to 0.75, which approximately corresponds to the distance where the van der Waals potential associated with two atoms begins increasing steeply. We denote the set of closed clash-

free conformations of  $L_B$  by  $\mathbf{Q}_{\text{closed}}^{\text{free}}$ . It has the same dimensionality as  $\mathbf{Q}_{\text{closed}}$ , but its volume is usually a small fraction of that of  $\mathbf{Q}_{\text{closed}}$ .

## References

Coutsias, E.A., Soek, C., Jacobson, M.P., and Dill, K.A. A kinematic view of loop closure, *Journal of Computational Chemistry*, **25**:510-528, 2004.

Liu, G., Milgram, J., Dhanik, A., and Latombe, J.C. On the inverse kinematics of a fragment of protein backbone. *Proc. 10th Symp. on Advances in Robot Kinematics (ARK)*, Ljubljana, Slovenia, June 2006.